Journal of Bioscience & Bioengineering

Gloriosa Superba Tissue Cultured Plant: Effect Of Abiotic Stresses On The Adaptation Of Metabolites Like Total Phenols, Flavonoids, And Alkaloids

Dharmendra Singh Khichi, A. S. Yadav

Department of Botany, Government. M.V.M., Bhopal, India.

Corresponding Author: Dharmendra Singh Khichi, A. S. Yadav, Department of Botany,Government. M.V.M., Bhopal, India.

Received Date: Dec 05, 2022 Accepted Date: Dec 08, 2022 Published Date: Jan 09, 2023

ABSTRACT:

Cultured G. superba plants on Murashige-Skoog (MS) medium were subjected to abiotic stress created artificially by applying different light periods (12, 14 and 16 hrs), different pH (6, 7 and 8.5), and different temperatures (25, 30 and 35°c) in order to study the effect of stresses on secondary metabolite adaptation. To ascertain the impact of abiotic stress on the quantity of phenol, flavonoids, and alkaloids, G. superba extract, extracted with 95% methanol, was utilised. The amount of phenolics ranged from 0.4440.004 to 1.3920.02 mg TA/g extract, and there were no appreciable differences between the plants grown under various physical conditions.

The highest TPC values were reported to be 1.1760.007, 1.3010.02 mg TA/g extract, and 1.3920.02 mg TA/g extract in plants that were cultivated at various temperatures. and 35 degrees Celsius, respectively. Plant extracts contained flavonoids at levels ranging from 0.0320.03 to 0.2680.002 mg QE/g extract. The extracts of G. Superba cultivated under temperature stress at 25, 30, and 35oC contained the largest amounts of flavonoids, 0.241, 0.268, and 0.247 mg QE/g extract, respectively. Plants cultivated at a temperature of 25oC had a high alkaloids content (11.3080.41mg Col/g extract), but alkaloids accumulated less slowly at higher temperatures (30 and 35oC). In contrast to pH and photoperiods, the present study found that temperature was significantly more important for the optimal adaptation of secondary metabolites in G. superba.

Keywords:

Gloriosa superba, Abiotic stress, Alkaloids, Flavonoids, Phenols;

INTRODUCTION :

Stress is a physiologically changed condition brought on by things that

tend to upset the balance. Any physical or chemical change brought on by a stress is referred to as strain [1]. The term "stress" has several diverse definitions, including the physiological definition and the suitable term as a response to certain circumstances. One of the most successful methods for increasing the output of bioactive secondary metabolites is the use of elicitors of plant defence systems, or elicitation [2]. In order to speed up the production of high product concentrations, secondary metabolite creation is stimulated in plant cell cultures using biotic and abiotic stressors that are categorised according to their source. Numerous environmental stressors, including extreme heat and cold, drought, alkalinity, salinity, UV radiation, and pathogen infection, have the potential to be hazardous the vegetation. Elicitation has frequently been used to boost production of secondary metabolites or to cause their denovo synthesis.

Plant cell cultures grown in vitro [3]. Several scientists have used a variety of conditions to increase the synthesis of secondary metabolites in cultures of plant cell, tissue, and organ. Phenylpropanoids are frequently accumulated more as a result of environmental challenges such pathogen invasion, UV radiation, lighting, wounding, nutrition deficiency, temperature, and herbicide application. A plant that is more resistant might be produced by secondary metabolite concentrations that are higher. They are believed to be expensive to produce and hinder plant growth and reproduction [4].

Different plants create secondary metabolites, a type of bioactive molecules that include numerous groups of organic compounds such alkaloids, terpenoids, phenols, flavonoids, tannins, saponins, etc. These chemical compounds in plants that have distinct physiological effects on the human body are what give them their therapeutic worth.[5] Phytochemical study of traditional medicines Because of its importance for finding new sources of bioactive chemicals and medicinal agents, research on secondary metabolites is a crucial area of basic science [6]. G. superba L., a perennial tuberous climbing herb that belongs to the Liliaceae family, is found across India's tropical and subtropical regions. One of today's most significant medicinal plants, G. superba L., is under danger of local extinction owing to climate change. The plant's various parts have a wide range of uses, particularly in Indian traditional medicine that dates back thousands of years. G. superba L. tubers and seeds are a pricey export product. This plant was harvested from the wild and utilised as medicine because of its medical potential.

95% of the medicinal plants used as raw materials for large-scale pharmaceutical businesses were out to be endangered species that were included to the Red Data Book due to over-exploitation [7]. This is why the current study is intended to look at how stresses affect the adaptability

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of secondary metabolites including phenol, flavonoid, and alkaloids in the G. superba tissue culture plant.

The Existing Ranking Methods:

In order to accomplish this, cultivated plants in Murashige-Skoog (MS) medium were subjected to abiotic stress that was induced artificially by applying varied light periods (12, 14 and 16 hrs), different pH (6, 7 and 8.5), and different temperatures (25, 30 and 35°c). Cultured plants were examined on the medium for a month while stressful conditions persisted. All plants were taken after one month in order to examine how abiotic stress affected the levels of secondary metabolites.

Extraction and Phytochemical Analysis:

At room temperature (39 °C), the cultured entire plants were dried and ground into powder. G. superba dry powder that was extracted using a soxhlet device and 95% methanol (Merck). To get crude for phytochemical analysis, the extract was then evaporated in a water bath at 50°C [8].

Determination of Total Phenolic Content:

The method of Singleton and Rossi [9] was slightly modified in order to measure the total phenolic content (TPC) of the crude extracts of the G. superba plant. The total phenol was determined using the Folin Ciocalteau reagent in this technique (1:10 v/v diluted with distilled water). Following incubation, the blue color's development was seen. Then, using a spectrophotometer, the absorbance of blue colour in several samples was determined at 725 nm. On the basis of a standard curve for tannic acid, the phenolic content was estimated as Tannic acid equivalents (TA)/g extract. Tannic acid equivalents (TA/g) of the plant extract were used to express the findings.

CONCLUSION:

The current study came to the conclusion that abiotic stress factors affect the plant's ability to produce secondary metabolites. The impacts are obvious. In actuality, the altered stress element affects productivity. For instance, the effects of temperature, pH, and photoperiod, among other factors, also affect plant productivity and adaptation.

There have been significant advancements in the generation of medicines and secondary metabolites using in vitro plant cell culture. The fundamental knowledge for the generation of secondary metabolites for commercial application will be provided through the utilisation of abiotic stress and changing environmental circumstances. Large-scale plant cell culture technology is now again gaining popularity because of the rising use of natural goods for medical purposes, low product yields, and supply issues associated with plant harvest. variables that are biotic and abiotic that affect secondary The potential to overproduce beneficial phytochemicals for a variety of uses is enhanced by metabolite synthesis. In this study, temperature exhibits the highest adaptability of secondary metabolites in G. superba plant as compared to pH and photoperiods. According to our study, G. superba's culture conditions were improved in order to produce bioactive secondary metabolite compounds.

REFERENCES:

- Gaspar, T.T., Franck, B., Bisbis, C., Kevers, L., Jouve, J.F., Hausman and Dommes., Concepts in plant stress physiology: Application to plant tissue cultures. Plant Growth Regul., 2002, 37, 263–285.
- Roberts, S. and Shuler, M.L., Large scale plant cell culture. Curr. Opin. Biotech., 1997, 8, 154-159.
- Barz, W., Danie, S., Hinderer, W., Jaques, U.,Kessmann, H., Koster, J. and Tiemann, K.,Elicitation and metabolism of phytoalexins in plant cell cultures. In: Pais, M., Mavituna, F.,Novais, J, editors. "Plant Cell Biotechnology".NATO ASI Series. Berlin: Springer-Verlag, 1988,211-230.
- Eilert, U., Elicitation: methodology and aspects of application. In: Constabel, F., Vasil, I., editors. "Cell Culture and Somatic Cell Genetics of Plants". 4, San Diego: Academic Press, 1987,153-196.
- Dicosmo, F. and Tallevi, S.G., Plant cell cultures and microbial insult: interactions with biotechnological potential. Trends in Biotech., 1985, 3, 110-111.
- Dutta, B., Study of secondary metabolites of Gomphostemma niveum Hook. f. in Assam.India. J. Med. Plants Stud., 2014, 2(5): 24-28.
- Akinmoladun, A.C., Ibukun, A.C., Afor, E., Obuotor, E.M. and Farombi, E.O., Phytochemical constituent and antioxidant activity of extract from the leaves of Ocimum gratissimum. Sci. Res.Essays, 2007, 2(5):163-166.
- Singh, D., Mishra, M. and Yadav, A.S., Effect of growth regulators on micro propagation of Gloriosa superba L. from seeds and their acclimatization. Annu. Res. Rev. Biol., 2015,7(2): 84-90.
- Singleton, V. and Rossi, J., Colorimetry of total phenolic with phosphor molibdic phosphotungstic acid reagents. Am. J. Enol. Vitic., 1965, 16:144–158.
- Samatha, T., Shyamsundarachary, R., Srinivas, P.and Swamy, N.A., Quantification of total phenolic and total flavonoid contents in extracts of Oroxylum indicum L. Asian J.Pharm. Clin. Res., 2012, 5Suppl 4, 177-179.
- John, B., Sulaiman, C.T., George, S. and Reddy, V.R.K., Spectro photometric estimation of total alkaloids in selected Justicia species. Int. J.Pharm. Pharm. Sci., 2014, 6(5): 647-648.

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- 12. Nacif de Abreu and Mazzafera., Effect of water and temperature stress on the content of active constituents of Hypericum brasiliense choisy.Plant Physiol. Bioch., 2005, 43(3):241-248.
- Boo, H., Heo, B., Gorinsteinc, S., Chond, S.U., Positive effects of temperature and growth conditions on enzymatic and antioxidant status in lettuce plants. Plant Sci., 2011, 181:479–484.
- Zahir, A., Abbasi, B.H., Adil, M., Anjum, S., Muhammad, Zia, & Ihsan-ul-haq., Synergistic Effects of Drought Stress and Photoperiods on Phenology and Secondary Metabolism of Silybum marianum. Appl. Biochem. Biotechnol., 2014, 174:693–707.
- Rangel, S., Abioti c stress based bioprocesses for the production of high value antioxidant phenolic compound in plants: An Overview. Revista Mexicana de Ingenieria Quimica, 2014, 13(1): 49-61.
- Swieca, M., Elicitation with abiotic stresses improves pro-health constituents, antioxidant potential and nutritional quality of lentil sprouts. Saudi J. Biol. Sci., 2015, 22(4): 409–416.
- Sawada, H., Shim, I.S., and Usui, K., Induction of benzoic acid 2-hydroxylase and salicylic acid biosynthesis modulation by salt stress in rice seedlings. Plant Sci., 2006, 171: 263-270.
- Singha, A., Lawrencea, K., Pandita, S. and Lawrenceb, R.S.,. Response of leaves, stems and roots of Withania somnifera to copper stress. Int.J. Pl. An. Env. Sci., 2014, 4(3): 60-67.
- Abu Bakar, M.F., Mohamed, M., Rahmat, A. and Fry, J., Phytochemicals and antioxidant activity of different parts of bambangan (Mangifera pajang) and tarap (Artocarpus odoratissimus). Food Chem., 2009, 113: 479–483.

- Grace, S.C. and Logan, B.A., Energy dissipation and radical scavenging by the plant phenylpropanoid pathway, Philosophical Transactions of the Royaln Society of London, 2000, 355: 1499-1510.
- Chutipaijit, S., Cha-Um, S. and Sompornpailin,K., Differential accumulations of proline and flavonoids in indica rice varieties against salinity.Pak. J. Bot., 2009, 41(5): 2497-2506.
- 22. Yamamoto, N., Mori, K., Numata, M., Koyama, K. and Kitayama, M., Effects of temperature and water regimes on flavonoid contents and composition in the skin of red-wine grapes. J. Int.Sci. Vigne. Vin., special issue Macrowine, 2010,13(29): 75-80.
- Pan, J., Vicente, A.R., Martinez, G.A., Chaves, A.R. and Civello, P.M., Combined use of UV-C irradiation and heat treatment to improve postharvest life of strawberry fruit. J. Sci. Food.Agr., 2004, 84: 1831-1838.
- Zengqiang, M.A., Shishang, Li., Meijun, Z. and Wojtaszek, P., Oxidative stress, antioxidant tolerance. Trends of Plant Sci. 2002, 9: 405–410.
- Hajiboland, R., Bastani, S. and Rad, S.B., Effect of light intensity on photosynthesis and antioxidant defense in boron deficient tea plants. Acta Biologica Szegediensis, 2011, 55(2):265-272.
- Toivonen, L., Laakso, S., Rosenqvist, H., The effect of temperature on growth, indole alkaloid accumulation and lipid composition of Catharanthus roseus cell suspension cultures.Plant Cell Reports, 1992, 11(8): 390-394.
- 27. Mua, H., Wang, R., Li, X., Jiang, Y.M., Wang, C.Y., Quan, Y.P., Peng, F. and Xia, B., Effect of Abiotic and Biotic Elicitors on Growth and Alkaloid Accumulation of Lycoris chinensis Seedlings. Z. Naturforsch, 2009, 64: 541 – 550.